

Evaluation of marine zooplankton community structure through environmental DNA metabarcoding

Anni Djurhuus¹, Kathleen Pitz², Natalie Sawaya¹, Jaimie Rojas-Marqu ez³, Brianna Michaud¹, Enrique Montes¹, Frank Muller-Karger¹, and Mya Breitbart¹
Limnology and Oceanography Methods: doi: 10.1002/lom3.10237

1. College of Marine Science, University of South Florida, FL, USA.
2. Monterey Bay Aquatic Research Institute, Monterey, California, USA.
3. Fundaci n La Salle de Ciencias Naturales, Estaci n de Investigaciones Marinas, Isla de Margarita, Venezuela.
Contact: Dr. Anni Djurhuus: Email: anni.djurhuus@gmail.com. Twitter: @AnniDjurhuus



Introduction

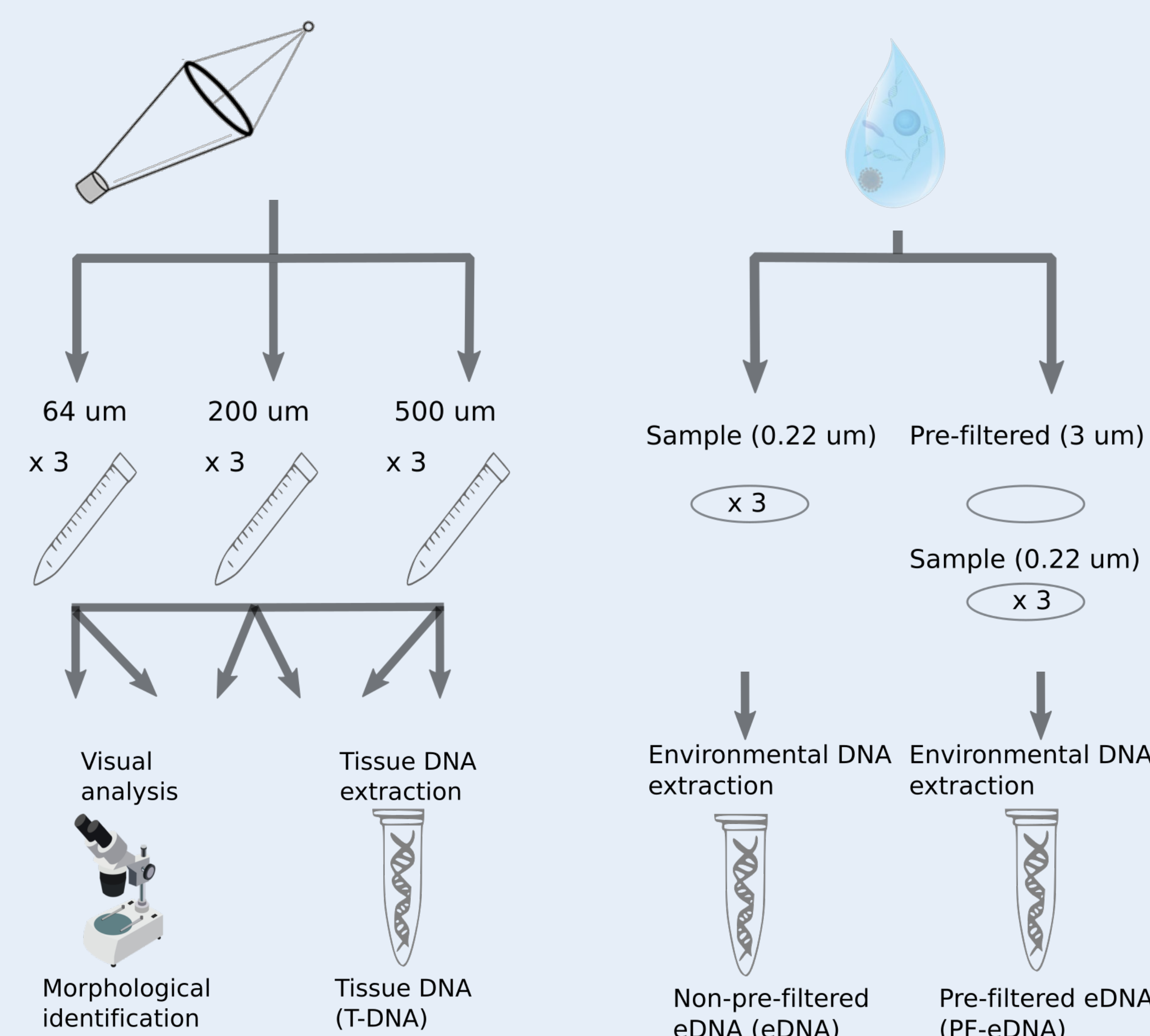
An essential element of environmental conservation programs is biodiversity assessment. Traditional methods that characterize biodiversity are laborious and can be environmentally destructive (1). Genetic analysis of environmental DNA (eDNA), which contains DNA shed by organisms present in a given environment, offers a cheaper, more sensitive, and less destructive method for characterizing biodiversity (2).

Objective

Here we aim to assess methods for comparing net tow tissue sequencing (T-DNA) with eDNA for two different genetic markers (18S and COI) and comparing those to morphological assessment of net tow samples.

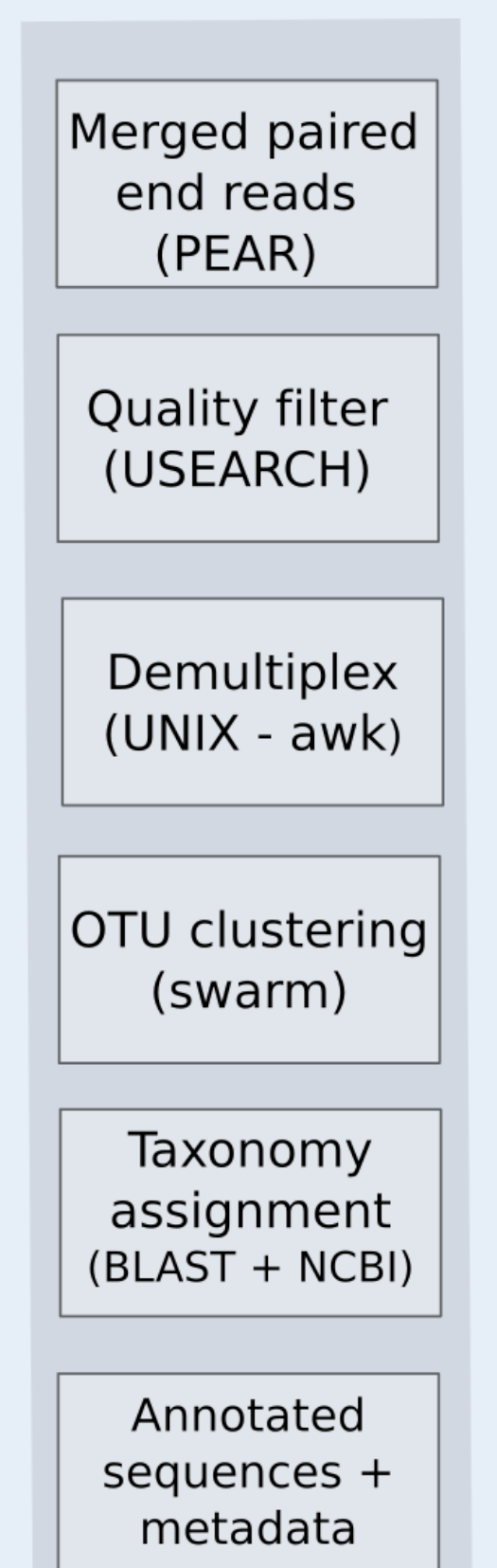
Methods

Seawater samples were collected onboard the R/V Walton Smith in May 2016 using a Conductivity Temperature Depth sensor fitted on a rosette with 12 Niskin bottles for water sampling. See Fig. 1 for laboratory and data workflow.



PCR and MiSeq sequencing (18S rRNA and COI) (Amaral-Zettler et al. 2009, Folmer et al. 1994; Leray et al. 2013 (refs. 3-5))

Data analysis



Results

- The two genetic loci (COI and 18S rRNA) recover different organisms on a Class level (i.e. chordates: ray-finned fishes (Class Actinopteri) and ascidians (Class Ascidiacea)) (Fig. 2).
- The tissue DNA (T-DNA) resembled the zooplankton taxonomic composition identified by microscopy more closely than the environmental DNA (eDNA) or pre-filtered environmental DNA (PF-eDNA).
- The morphologically identified taxa were more similar to the 18S rRNA taxonomy than to the COI.
- Both genes detected the same taxonomic groups but at different abundances (Fig. 2-3).
- The most abundant copepod genera were detected with all treatments (Fig. 3).

Figure 1 Schematic of sample collection and processing pipeline. Pre-filtered eDNA (PF-eDNA), eDNA, and tissue DNA (T-DNA).

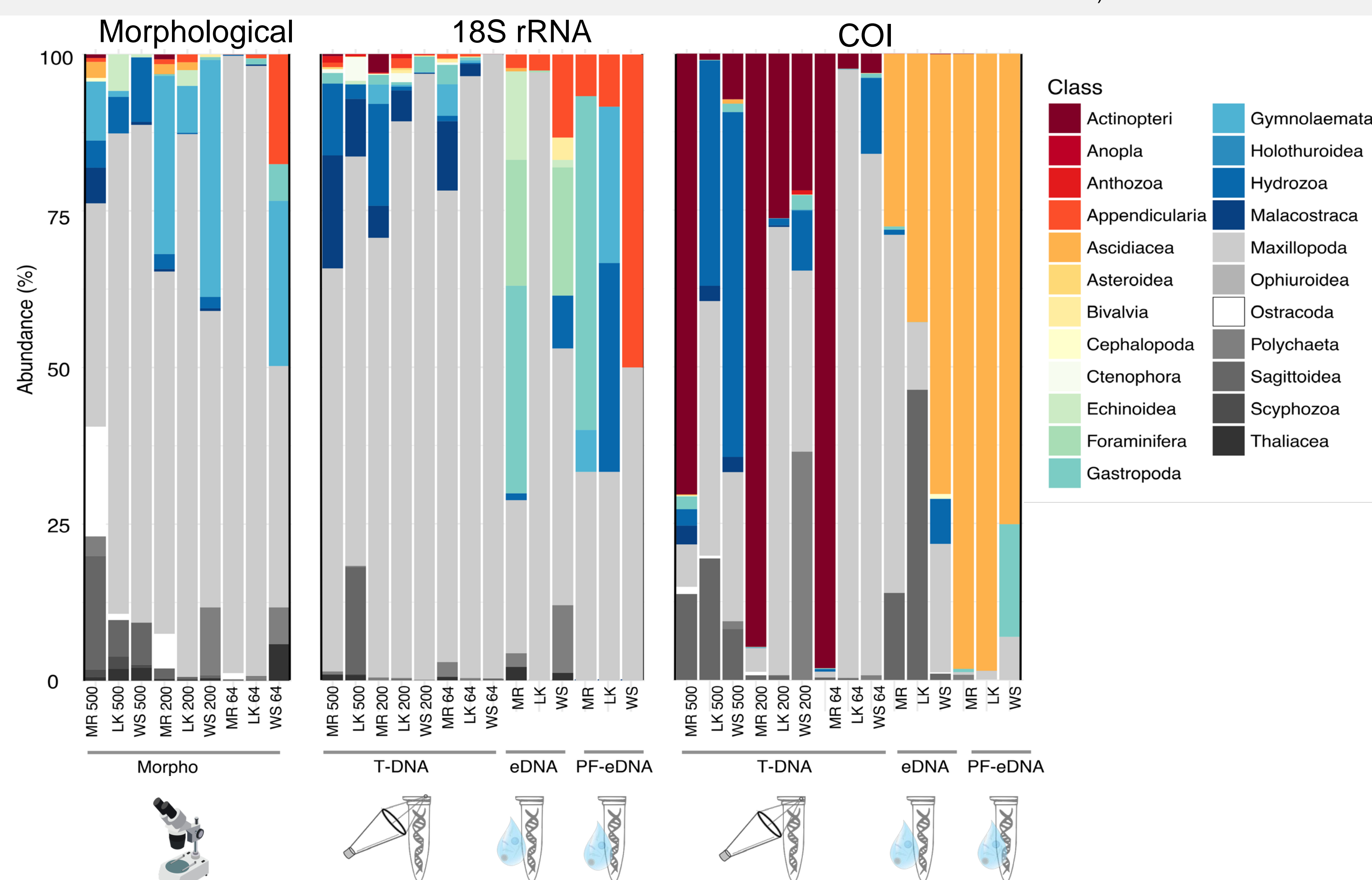


Figure 2 Barplot of all data at the Class level. The numbers on the labels for each bar refer to the mesh size (in μm) used in the net tows, and letters represent the sampling stations (Molasses Reef (MR), Looe Key (LK), and Western Sambo (WS)).

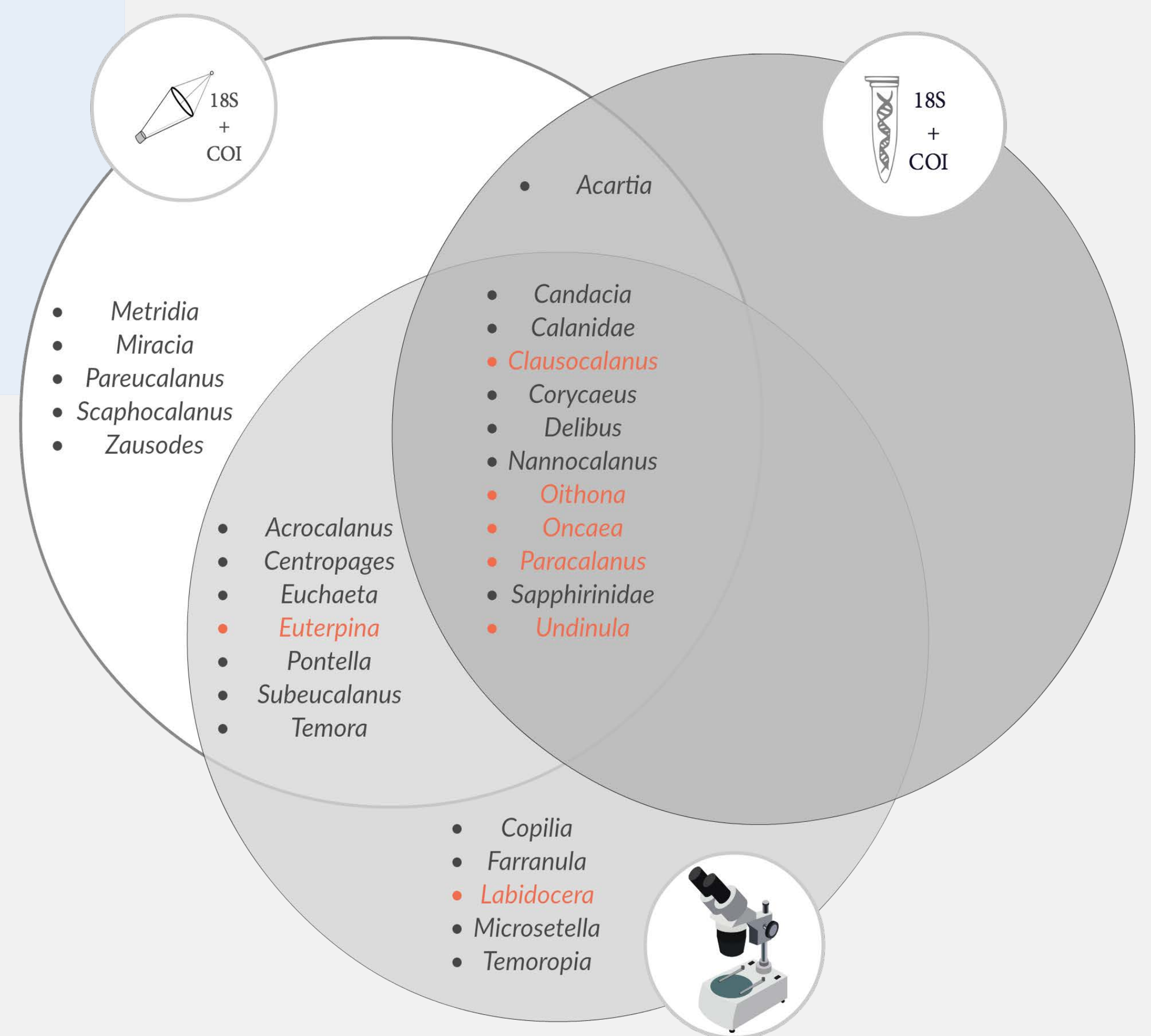


Figure 3. Venn diagram of copepod genera detected using genetic markers in tissue DNA (T-DNA, left), environmental DNA (eDNA, right), and morphological assessments (bottom). Nearly all dominant (red, >5% of total abundance) copepod genera were identified by each treatment.

Acknowledgements

This work was supported by NASA grant NNX14AP62A 'National Marine Sanctuaries as Sentinel Sites for a Demonstration Marine Biodiversity Observation Network (MBON)' funded under the National Ocean Partnership Program (NOPP RFP NOAA-NOS-IOOS-2014-2003803 in partnership between NOAA, BOEM, and NASA).

References

- Wheeler et al. 2004, Graellsia 61: 151-160.
- Harvey et al. 2017, Journal of Exp. Mar. Bio. and Eco. 487, 113-126.
- Amaral-Zettler et al. 2009, PLoS ONE 4(7): e6372.
- Folmer et al. 1994, Mol. Biol. Biotechnol. 3:294-299.